Attorney D cket No: 20200/2092 (Serial No.:09/889,802)

 Inventor: Kreutzer, et al. Response and Amendment

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under 35 U.S.C. § 365 to PCT/DE00/00244, filed January 29, 2000, all of which are incorporated by reference herein.

Please replace the paragraph at page 1, lines 4-7, with the following replacement paragraph:

Background of the Invention

This invention relates to double-stranded ribonucleic acid (dsRNA), its use in mediating RNA interference in vitro and in vivo, and compositions and cells comprising the dsRNA.

Please replace the paragraph at page 2, lines 31-33, with the following replacement paragraphs:

-- Summary of the Invention

The invention relates to an oligoribonucleotide having a double stranded structure (dsRNA). The oligoribonucleotide comprises two separate RNA strands, wherein one strand of the dsRNA has a region which is complementary to an RNA transcript of at least a part of a target gene, wherein the region is not more than 49 nucleotides in length, and wherein the target gene is a mammalian gene. The oligoribonucleotide may have a length of between 15 and 49 base pairs, and the RNA transcript may be a primary or a processed RNA. The oligoribonucleotide may comprise a linker between the two RNA strands, such as a polyethylene glycol linker. The oligoribonucleotide may be modified so as to be resistant to RNA degradation. The oligoribonucleotide may comprise a 3' overhang, such as a single nucleotide overhang. The oligoribonucleotide may be 21 or 23 nucleotides in length.

In another aspect, the invention relates to a method for inhibiting the expression of a target gene in a mammalian cell, such as a human cell. The method comprises (a) introducing into the cell an oligoribonucleotide having a double stranded structure (dsRNA), comprising two separate RNA strands, wherein one strand of the dsRNA has a region which is complementary to an RNA transcript of at least a part of a target gene; wherein the region is not more than 49 nucleotides in length; and (b) maintaining the cell produced in step (a) for a time sufficient to obtain degradation of an RNA transcript of the target gene, thereby inhibiting expression of the



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target gene in the cell. The oligoribonucleotide may have a length of between 15 and 49 base pairs, and the RNA transcript may be a primary or a processed RNA. The oligoribonucleotide may comprise a linker between the two RNA strands, such as a polyethylene glycol linker. The oligoribonucleotide may be modified so as to be resistant to RNA degradation. The oligoribonucleotide may comprise a 3' overhang, such as a single nucleotide overhang. The oligoribonucleotide may be 21 or 23 nucleotides in length.

In yet another aspect, the invention relates to a mammalian cell comprising an exogenous oligoribonucleotide, wherein the oligoribonucleotide has a double stranded structure (dsRNA) comprising two separate RNA strands, and wherein one strand of the dsRNA has a region which is complementary to an RNA transcript of at least a part of a target gene. The mammalian cell may be a human cell, the region may have not more than 49 nucleotides in length, the oligoribonucleotide may have a length of between 15 and 49 base pairs, and the RNA transcript may be a primary or a processed RNA. The oligoribonucleotide may comprise a linker between the two RNA strands, such as a polyethylene glycol linker. The oligoribonucleotide may be modified so as to be resistant to RNA degradation. The oligoribonucleotide may comprise a 3' overhang, such as a single nucleotide overhang.

In still another aspect, the invention relates to a composition comprising an oligoribonucleotide as described above. The composition may further comprise a second oligoribonucleotide, wherein the second oligoribonucleotide differs in sequence from the oligoribonucleotide.

Please add the following paragraphs and headings directly below the foregoing replacement paragraphs:

Brief Description of The Drawings

FIG. 1 is a schematic representation of a plasmid for the *in vitro* transcription with T7-and SP6-polymerase.

FIG. 2 shows RNA following electrophoresis on an 8% polyacrylamide gel and staining with ethidium bromide.

